Roles of Ions in Formulation used for Collaborative Treatment of Disease

Pratiksha B. Avhad, Dr. V. U. Barge, Kamlesh R. Deshmane

Department of Pharmaceutical Quality Assurance, PDEA Shankarrao Ursal College of Pharmaceutical Sciences and Research Centre, Kharadi, Pune, Maharashtra, India

ABSTRACT

Cancer is one of the problems of severe threats to human life and health. The aim of the present study was to examine the effects of calcium channel protein on cancer cells. Calcium is a versatile element that participates in cell signaling for a wide range of cell processes such as death, cell cycle, division, migration, invasion, metabolism, differentiation, autophagy, transcription, and others. Calcium channels may be generally categorized into two major classes: Voltage-gated calcium channels (VGCCs) and ligand-gated calcium channels (LGCCs). VGCCs may be classified into five subtypes: L type, N-Type, P-type, T-type and R-type. The immune system is specialized in the process of cancer cell recognition and elimination, and is regulated by different ion channels. The success of treatment depends upon the type of cancer, locality of tumor, and its stage of progression. Surgery, radiation-based surgical knives, chemotherapy, and radiotherapy are some of the traditional and most widely used treatment options. Some of the modern modalities include hormone-based therapy, anti-angiogenic modalities, stem cell therapies, and dendritic cell-based immunotherapy.

KEYWORDS: Ca²⁺channels, Ion channels, voltage-gated ions, cancer therapy, tumors

How to cite this paper: Pratiksha B. Avhad | Dr. V. U. Barge | Kamlesh R. Deshmane "Roles of Ions in Formulation used for Collaborative Treatment of

Disease" Published International Journal of Trend in Scientific Research and Development (ijtsrd), ISSN: 2456-6470, Volume-7 Issue-2, April 2023, pp.1000-1010,



www.ijtsrd.com/papers/ijtsrd56202.pdf

Copyright © 2023 by author (s) and International Journal of Trend in Scientific Research and Development

Journal. This is an Open Access article distributed under the



terms of the Creative Commons Attribution License (CC BY 4.0) (http://creativecommons.org/licenses/by/4.0)

INTRODUCTION

Calcium channels may be generally categorized into two major classes: Voltage-gated calcium channels (VGCCs) and ligand-gated calcium channels (LGCCs). VGCCs may be classified into five subtypes: L-type, N-Type, P-type, T-type and R-type. These ion channels have been implicated in the progression of numerous cancers. Ion channels are well recognized as important therapeutic targets for treating a number of different pathophysiologies. Cancer is one of the problems of severe threats to human life and health. Metal ions have a very important impact on life systems, and they play a necessary role that other chemical molecules cannot replace 1. Excess, deficiency, and abnormal distribution of metal ions will seriously affect various physiological properties of cells. Finding new and innovative treatments for cancer is a major problem across the world². Radiation therapy is one of the oldest modalities for cancer treatment and is currently prescribed to more than 50% of all patients. It is based on delivering high doses of ionizing radiation to well-localized tumor targets in the body. The goal

is to kill all the tumor cells with acceptable toxic effects to the surrounding normal tissue, which is unavoidably exposed. Indeed, radiotherapy success is limited by the toxicity in the normal tissue. Metal ions significantly impact the biosystem and play an essential role in diverse physiological activities such as maintaining cell homeostasis, regulating metabolic pathways, substance synthesis, signal transmission, and energy conversion³. Metal ions have previously been used in imaging technology, such as the contrast agents based on Gd3+, which have dominated the MRI field for several decades 4. However, the problems of traditional contrast agents limit their prospects for biological applications 5, such as insufficient time in vivo circulation of iodine-based and barium based agents ⁶ the short half-life period of radioactive 18F⁷ and the high toxicity of gadolinium ions ⁸. Ion channels are important drug targets because they play a crucial role in controlling a very wide spectrum of physiological processes ⁹ because their dysfunction can lead to pathophysiology ¹⁰. Given the strong historical precedent that exists for discovering and commercializing successful drugs that modulate the activity of voltage-gated sodium, calcium, or potassium channels, or ligand-gated ion channels, new generations of therapeutic agents are expected to result from targeting this protein family.

Ion Channels in Cancer:

Ion channels comprise an important factor influencing the formation and development of tumors. Such malignant transformation leads to enhanced proliferation, abnormal differentiation, impaired apoptosis, and finally uncontrolled migration and Ion channels and anti-cancer invasion (Table 1). This is often associated with altered levels of ion channel expression as well as their activity in the mutated cancer cells¹¹. The role of ion channels in pathogenesis of various diseases including cancer and its treatment has been extensively studied. The major types of ion channels implicated in carcinogenesis are presented below

Table 1: The role of distinct ion channels in cancer development and progression

Ion channels	Expression profile	Cancer type	References
Proliferation of cancer cells Shaker-like K+ channels (Kv1.1, Kv1.3, Kv1.5)	Gene and protein upregulation	Glioma, breast cancer, lung cancer, pancreas cancer, prostate cancer, lymphoma	12, 13
EAG K+ channels (EAG1, EAG2) Gene and protein upregulation	Gene and protein upregulation	Medulloblastoma, breast cancer, head and neck cancer, melanoma, gastrointestinal tract cancer	14-16
EAG-related K+ channels (HERG/Kv11.1)	Gene and protein upregulation	Melanoma, neuroblastoma, breast cancer	17
Ca2+-activated K+ channels (KCa3.1)0	Gene and protein upregulation	Glioma, breast cancer, lung cancer, pancreas cancer, prostate cancer, lymphoma	18-21
Cell migration and metastasis EAG K+ channels (EAG1/ Kv10.1)	Gene and protein upregulation	end in Scientific Migration of breast cancer cells levelopment	22
Ca2+-activated K+ channels	Gene and protein upregulation	Breast cancer → metastasis to brain Breast cancer → bone metastasis Migration of glioma cells, transformed renal epithelial cells and breast cancer cells	23-27
Tumor angiogenesis EAG K+ channels (EAG1	Gene and protein upregulation	Breast cancer and other solid tumors	

Ca2+ channels:

The intricate fluxion of Ca2+ ions between extracellular and intracellular stores shapes the movement of Ca2+, such as Ca2+ release, Ca2+ oscillations, and Ca2+ spikes, modulating numerous biological functions $^{28-29}$. It is not surprising that the exchange of Ca2+ ions among different components of cells is interconnected and highly coordinated, and uncontrolled remolding of this well-connected network may lead to cancer cells metastasis to bone. Extracellular Ca2+ concentration is maintained at a high level (~1–2 mmol/L), which is 10–20,000 times that of the cytosolic Ca2+ concentration (~100 nmol/L). Endoplasmic reticulum (ER) stores intracellular Ca2+ ions, with a Ca2+ concentration around 100–400 μ mol/L 30 . The regulation of this gradient is operated through a variety of mechanisms (Figure 1). Plasma membrane Ca2+ ATPases (PMCAs) and sarco(endo)plasmic reticular Ca2+ ATPases (SERCAs) are the main ATP-dependent channels that extrude Ca2+ ions from the cytosol to the extracellular space and ER, respectively. Inositol 1,4,5-trisphosphate receptors (IP3 Rs) initiate Ca2+ releasing from the ER 31 . After the depletion of the intracellular Ca2+ stores, store-operated Ca2+ entry (SOCE), a specific Ca2+ influx pathway, initiates Ca2+ influx through Orai1 Ca2+ channels after activation by the ER Ca2+ store sensor stromal interaction molecule 1 (STIM1 $^{32\cdot33}$. Extracellular Ca2+ ions enter the cytoplasm through substantial mechanisms and are the primary origin for intracellular Ca2+ signaling in cells. Examples include store-operated Ca2+ channels (SOCs), the transient receptor potential (TRP) superfamily of ion channels, voltage-gated Ca2+ channels (VGCCs) including L-, R-, N-, P/Q-, and T-type channels, and stretch-activated PIEZO channels $^{31,34\cdot37}$.

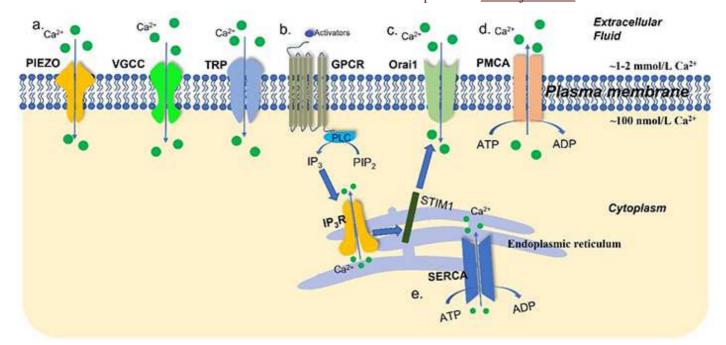


Figure 1. An overview of Ca2+ channels, transporters, and pumps in the plasma membrane and ER. Intracellular Ca2+ concentration is governed by a tightly mediated mechanism. (a) The TRP channels, VGCCs, and stretch-activated PIEZO channels are the Ca2+ channels and transporters in the plasma membrane; (b) after stimulation by activators, G-protein-coupled receptors (GPCRs) facilitate the dephosphorylation of phosphatidylinositol 4,5-bisphosphate (PIP2) into inositol 1,4,5-trisphosphate (IP3) by phospholipase C (PLC). In turn, IP3 Rs initiate Ca2+ release from the ER; (c) STIM1 senses the depletion of the ER Ca2+ stores and activates Ca2+ influx via Orai1 Ca2+ channels; (d) PMCAs extrude Ca2+ ions from intracellular space to the extracellular space; (e) SERCAs transport Ca2+ from the cytoplasm into ER. ADP: adenosine diphosphate

Specific calcium channels or pumps as targets for cancer therapy

The overexpression of specific calcium channels and pumps in some cancer types and/or subtypes has led to the proposal that pharmacological modulators of some calcium channels or pumps may represent future cancer therapies. However, overexpression itself is not a sufficient criterion for a potential pharmacological target in cancer. Pharmacological modulation of the target must alter proliferation, migration and/or induce cancer cell death, analogous to the way anti-HER2 agents exploit the overexpression of the HER2 protein in some breast cancers to control disease progression [38]. The critical role of the calcium signal in many of the hallmarks of cancer [39] certainly gives the potential for an effective therapy. However, a lack of activity, defective membrane trafficking or a limited role of a specific calcium pump or channel in a pathway relevant to tumor progression are just some of the examples where over-expression per se will be insufficient for a calcium pump or channel to be a therapeutic target in a particular cancer type. Another consideration is the likely effects of global pharmacological modulation of the target. Although many cancer therapies work on targets with critical roles in normal cells, as exemplified by some of the major side effects of some anticancer agents (e.g. immunosuppression), this is an important consideration. Tools for target selection and/or prioritization could include consideration of the known toxicity of pharmacological modulators to the target, or where such agents are not currently available, the viability and/or phenotype of knockout animals. It should be noted that despite the diverse expression of some ion channels and ion pumps they still represent targets highly amenable to drug development. Indeed, ion channels have been reported to represent 19% of human protein drug targets and are the targets of existing therapies including L-type Ca²⁺ channel blockers for the control of hypertension [40].

A number of reviews have highlighted the potential of calcium permeable ion channels and calcium pumps as therapeutic targets [41,42]. An example where this approach has recently started to extend towards human clinical trials is for the targeting of the highly Ca²⁺-selective ion channel TRPV6 [43]. Increased levels of TRPV6 have been reported in a variety of malignant cancers [44] including estrogen receptor negative breast cancers where TRPV6 overexpression may be driven by increases in TRPV6 copy number [45]. Silencing and overexpression studies have identified a critical role for TRPV6 in the proliferation of some cancer cell lines using in vitro and in vivo models [46,47]. TRPV6 inhibitors have now undergone human phase 1 clinical trials in patients with advanced tumors of epithelia origin including those of the ovary, colon, pancreas, breast and prostate [43]. Fig. 1 presents a conceptual overview of how an overexpressed ion channel could be

pharmacologically targeted for cancer therapy. The first approach is valid for ion channels that contribute to the proliferation and/or invasive pathways in cancer cells through effects on cytosolic Ca²⁺ signaling. For these targets pharmacological inhibition may attenuate these pathways to reduce proliferation and/or invasion. Example of this approach include the aforementioned TRPV6 as well as inhibitors of store operated calcium entry (SOCE) which target the remodeling of Orai1-mediated Ca²⁺ influx which appears to be a feature of some cancer types [48,49,50]. TRPV4 is another example, since TRPV4 silencing has recently been shown to reduce the invasiveness of breast cancer cells [51]. In the specific context of TRPV4, it is interesting to note the successful completion of a phase 2 clinical trial of the TRPV4 inhibitor GSK2798745 in patients with congestive heart failure [52], highlighting the potential for the repurposing of this compound and/or similar agents for the treatment of some cancers. Indeed, there is a variety of opportunities for the repurposing of agents targeting calcium permeable ion channels for cancer, such as T-type Ca²⁺ channel blockers such as mibefradil (previously used for cardiovascular disease) which is being assessed in clinical trials for glioblastoma multiforme [53], the most common and aggressive form of brain cancer. The focus of drug development programs for SOCE inhibitors (developed by companies such as CalciMedica and Rhizen Pharmaceuticals) for an array of conditions including autoimmune disorders [54] also represent opportunities for drug repurposing.

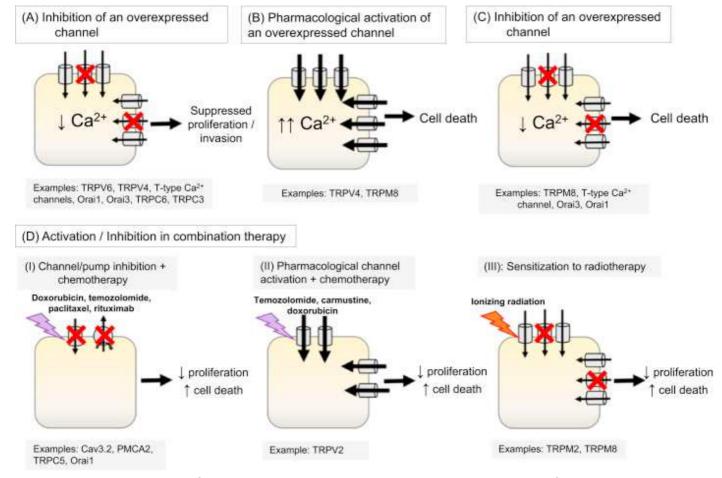


Fig. 1. Strategies to target Ca^{2+} signaling for cancer therapy. The overexpression of Ca^{2+} channels in cancer cells may be targeted by: (A) inhibition to suppress pro-proliferative or pro-migratory Ca^{2+} signals, examples of such channels include TRPV6 [47], T-type voltage-gated Ca^{2+} channels, i.e. Cav3.2 [55], Cav3.2 [56], TRPC6 [57] and TRPC3 [58]; (B) activation to promote Ca^{2+} overload and activating cell death (eg: apoptosis and oncosis), examples are TRPV4 [59] and TRPM8 [60]; or (C) inhibition to suppress pro-survival Ca^{2+} signals to induce cell death, examples include TRPM8 [60], T-type Ca^{2+} channels [61], Cav3.2 [62] and Cav3.2 [63]. Alternatively, Cav3.2 signaling can also be strategically targeted in combination with anti-cancer drugs to promote killing of cancer cells (D). Proposed mechanisms include (I) inhibition of a Cav3.2 [64], TRPC5 [65] and Cav3.2 [66] or a plasmalemmal Cav3.2 efflux pump such as PMCA2 [67]; (II) activation of a Cav3.2 channel, eg: TRPV2 [68] to promote the cytotoxic effect of chemotherapy drugs such as temozolomide, doxorubicin and carmustine; or (III) inhibition of specific Cav3.2 influx channels such as TRPM2 [69] and TRPM8 [70] in combination with radiotherapy to promote tumor-suppressing and/or cytotoxic effects.

Activation of Ca²⁺ permeable ion channels with pronounced overexpression represents another

approach to promote cancer cell death, and this potential has now been demonstrated in a variety of

models including TRPV4 in breast cancer cells [59] and TRPM8 in prostate cancer cells [60]. However it should be noted as reflected also in fig: 2, that there may also be examples where inhibition of a plasmalemmal Ca²⁺ permeable ion channel may also induce cancer cell death. In these examples rather than direct "Ca²⁺ overload" inducing necrotic or apoptotic cell death, the attenuation of a Ca²⁺ influx pathway may induce cancer death through other pathways. Examples includes the ability of T-type Ca²⁺ channel inhibition to produce apoptosis in p53competent HCT116 colon cancer cells through activation of p38-MAPK [61]. Another area of study has been the identification of calcium transporters, pumps and channels of intracellular organelles that when silenced can also attenuate pathways important in tumor progression. These examples extend to the recently identified components of mitochondrial Ca²⁺ transport and the less studied Ca²⁺ pumps of the Golgi apparatus – secretory pathway Ca²⁺ ATPases (SPCAs). Reduced expression of MCU in MDA-MB-231 breast cancer cells suppresses their migration in vitro and metastasis in vivo [71], and reduced expression of SPCA2 attenuates the proliferation of MCF-7 cells in vitro and tumor growth in vivo [72].

The discussion above has mostly focused on targeting specific calcium channels, pumps or exchangers through exploitation of their overexpression and/or critical role in a proliferation or invasive pathways. However, there may be circumstances where mutations in a Ca²⁺ regulating protein may be a feature of cancer cells. In some cases these events may be rare and confined to only a very few individual cancers, such as appears to be the case for gain of function mutations in Orai1 identified from the cBioPortal database [73]. Although further work on other calcium channels, pumps and exchangers is required, there is an example which does point to the potential significance of a mutation of a calcium pump in a cancer which leads to a significant clinical impact. Somatic mutations in the PMCA3 Ca²⁺ efflux pump in some aldosterone-producing adenomas reduces the activity of the PMCA3 pump and remodels Ca²⁺ signaling which appears to promote aldosterone production and the associated severe arterial hypertension in patients with these adenomas [74,75]. Whether the future will see highly selective agents targeting specific mutations of a calcium channel, pump or channel in a specific cancer type is still unclear, however such agents are theoretically possible given the successful development of CFTR ion channel mutation specific drugs for cystic fibrosis therapy [76].

Voltage-gated ion channels, cancer development, and metastasis

VGICs, in general, significantly contribute to a variety of mechanisms involved in cell survival and are crucial for maintaining normal tissue homeostasis, such as cell proliferation (77) cell migration (78), gene expression (79), vesicular patterning (80), apoptosis (81), and more.

All these mechanisms are critically important in maintaining and promoting cell activities but are also part of cancer cells proliferation. Increasing evidence supports ion channels in cancer cells *in vitro* and *in vivo*, revealing how they contribute to different aspects and stages of the cancer process.

According to their expression levels, several VGICs have been found to play essential roles during the cell cycle. Thus, aberrant ion channels' expression or malfunction can impair these processes, driving the transformation of normal cells into malignant ones that exhibit uncontrolled multiplication and spreading.

The following are a couple of examples:

Potassium channels: With 77 genes coding and many splice variants, the potassium channels are the largest, most diverse group of ion channels in the human genome. Voltage-gated potassium (Kv) channels play a pivotal role in the progression of various cancer types, including blood cancers such as leukaemia and lymphoma. Several studies have demonstrated an altered expression of the potassium channels subunits in cancer compared to normal tissues. However, the changes depend on the type and the stage of the disease.

In breast cancer, a significantly up-regulated expression of Kv1.3 channel mRNA is already observed in the first stage of the disease (82). Similarly, the human voltage-gated potassium channel ether à go-go 1 (EAG1, Ky 10.1) is overexpressed in most types of tumors, including leukaemia (83).

In the case of prostate cancer, instead, there is a significant inverse correlation between the expression of the Kv1.3 channels in the epithelium of human prostate tissue and the grade of the tumor (84).

Sodium channels: in a study by Fraser shows that, two rat prostate cancer cell lines with different metastatic abilities such as MAT-LyLu (strongly metastatic cell line expressing functional sodium channels) and AT-2 (weakly metastatic cell line with no functional sodium channels), were used in a comparative approach. The results plainly show that only the MAT-LyLu cells with functional VGSC

expression have enhanced prostate cancer cells' cellular motility (hence metastatic process) (85).

Another example is the expression of the sodium Nav1.5 ion channel in breast cancer. A study by Nelson shows that the Nav1.5 α subunit regulates breast tumor growth and potentiates migration and invasion, supporting the notion that compounds targeting Nav1.5 may help reduce metastasis (86).

A similar example is a study by Fraser showing that Nav1.5 expression is significantly up-regulated in metastatic human breast cancer cells and tissues compared with matched normal breast tissue and that Nav1.5 activity potentiates cellular directional motility, endocytosis, and invasion (87).

Other isoforms of Nav sodium channels such as Nav1.6 and Nav1.7 are involved in cervical cancer, breast, prostate, and non-small cell lung cancers.

Calcium channels: Calcium ion channels also have confirmed roles in cellular functions, including mitogenesis, proliferation, differentiation, apoptosis, and metastasis. For example, the expression of several calcium channels of the TRP superfamily is elevated in different common carcinomas, such as the TRPC1 in breast cancer, TRPC3 in some breast and ovarian cancer, TRPC6 in breast, liver, stomach, and glia cancers, and TRPM7 in breast, pancreas, ovarian in and gastric cancers, to mention some.

The high voltage-activated Cav1.2 channel is lop2. Classification by grade overexpressed in most cancer types, including colorectal, gastric, leukaemia, brain, uterus, breast, pancreatic, sarcoma, skin, and prostate. Similarly, the Cav1.3 is highly expressed in most types of cancer, including breast and prostate cancer, brain cancer, colorectal, gastric, bladder, lung, oesophageal, and uterine tumors (88,89). Also, in breast cancer, a total 5 VGCC family members (CACNA1A, CACNA1B, CACNA1E, CACNA1G, and CACNA1I) show a reduced expression.

Tumor biology

Cell division, when grows independent of growth factors, forms tumors, which involve a series of steps. In the very first stage, a large mass of cells known as hyperplasia is formed because of uncontrolled cell division. This is followed by dysplasia in which cell growth is accompanied with abnormalities. Additional changes occur in the next stage when these atypical cells start to spread over a limited area of the tissue, losing their original function. This phase is coined as anaplasia. At this stage, the tumor is not invasive and is considered as benign. In the advanced stage, the tumor cells acquire the ability to metastasize. They begin to invade the surrounding tissues as well as those located away via bloodstream.

This stage is considered to be malignant and is very hard to treat. However, not all tumors progress to this level, if identified earlier [14]. Though tumor cells are able to proliferate independent of growth factors, they still require nutrients and oxygen for their growth. All normal tissues are sufficiently supplied with capillaries for the supply of nutrients and oxygen to every cell. Similarly, tumors, as growth progresses, form new blood vessels in a process called as angiogenesis so that nutrients reach the cells located at the center of the tumor mass which do have access to normal blood vessels [15].

The types of tumor

- 1. On the basis of the type of cell initially altered Tumors are named depending upon the type of cell from which they originate. These include:
- > Carcinomas, which result from altered epithelial cells. They constitute the highest ratio in all types of cancer.
- Sarcomas denote the cancer abnormalities in the bone, muscle, fats, and connective tissue.
- Leukemia, which originate from cancerous white blood cells.
- Lymphoma, which is a malignancy of the lymphatic system or cells which are derived from the bone marrow (BM).
- Myelomas depict the cancers of those particular white blood cells that synthesize antibodies [14].

This is the abnormality in cells with respect to their surrounding normal tissues. Increase in abnormality increases the grade, from 1 to 4. Well-differentiated cells closely resemble normal cells and belong to lowgrade tumors. Improperly differentiated cells are highly abnormal with respect to the surrounding tissues [16]. These are high-grade tumors.

- Grade 1 : This includes well-differentiated cells having slight abnormality.
- : These cells are moderately differentiated Grade 2 and a bit more abnormal.
- : The cells are improperly differentiated Grade 3 and very abnormal in context of having mutated chromosomes and produce some harmful chemicals which affect nearby cells and may enter in the blood.
- Grade 4 : Cells are immature, primitive, and undifferentiated

3. Causes of cancer

Origin and advancement of cancer depend on many factors inside the cell (mutations, immune conditions, and hormones) as well as external factors from the environment (smoking, chemicals. organism, and radiations). These entire elements act together to cause abnormal cell behavior and uncontrolled proliferation. The resultant unusual cell mass in the body grows and affects normal tissues in their surroundings, and sometimes it also spreads to the other localities in the body (metastasis) [17] (Figure 1). According to the most accepted model for cancer causation, mutations in tumor suppressor and oncogenes is the major factor leading to the cancer development. Another model suggests that some mutation in a master gene that control the division of cells can also shepherd normal cells toward abnormal chromosomal replication, which can result in duplication or deletion of the entire sections of chromosomes [18]. This change in genetic content in the cells produces abnormal amount of a specific protein irrespective of the actual need. If any chromosomal aberration affects a protein that plays a crucial role in cell cycle, quantitatively or qualitatively, it may result in cancer. There is also a strong indication that the unnecessary addition (hypermethylation) or deletion (hypomethylation) of methyl groups to genes involved in the regulation of cell cycle, DNA repair, and apoptosis is also associated with some cancers. It is necessary to commemorate that cancers can take months to years for accretion of DNA mutations enough for the resultant cancer mass to be detectable. Thus, there can be several mechanisms which lead to the development of cancer. This further obscures the arch difficult task of defining the actual cause of cancer lop[3] Dow JAT. The essential roles of metal ions in [19].

> Mutations in the p53 tumor suppressor gene

Considering biochemical pathways the most important component central to human carcinogenesis is the P53 gene whose normal function is associated with gene transcription, DNA synthesis, apoptosis, and DNA repair [20]. Alterations and mutations in p53 elicit the development of primary tumors. The biochemical processes related to the normal function of p53 gene are performed by multiunit protein machines. The functions of these machines are altered by some viral oncoproteins, which bind with the p53 and perturb its interactions with other cellular protein components [21].

Linking tumor viruses to human cancer

Development of human malignancies is strongly associated with viruses. In fact, 15% of the cancer are believed to be caused by oncogenic viruses which include human papillomaviruses (HPVs), Epstein-Barr virus (EBV), Kaposi's sarcoma-associated herpes virus (KSHV, also known as HHV-8), and hepatitis B and C virus (HBV and HCV) [22]. Another virus known as Merkel cell polyomavirus (MCPyV) has been recently described causing Merkel cell carcinoma, a rare but aggressive type of skin

cancer [23]. The recent studies on these cancercausing agents have been very helpful to understand the basic biology of cell and how disturbances in the cellular pathways lead to the initiation and maintenance of cancer.

RESULT AND DISCUSSION:

It is clearly evident in the literature, that Ca²⁺permeable channels, transporters and pumps play important roles in a wide range of cancer-related process.

ACKNOWLEDGEMENT:

The authors would like to acknowledge PDEA's Shankarrao Ursal College of Pharmaceutical Sciences and Research Centre, Pune, Maharashtra, India for offering all vital help to accomplish this work efficaciously.

CONFLICT OF INTEREST:

All authors declared no conflict of interest for the work.

References:

- C. Andreini, I. Bertin, G. Cavallaro, G. L. Holliday and J. M. Thornton, J. Biol. Inorg Chem., 2008, 13, 1205.
- [2] Siegel, R. L., Miller, K. D., and Jemal, A. (2020). Cancer statistics, 2020. CA Cancer J. Clin. 70, 7–30. doi:10.3322/caac.21590.
- insect homeostasis and physiology. Curr Opin 2456-6470 Insect Sci. 2017; 23:43–50.
 - Scott PJH. A picture is worth a thousand words: the power of neuroimaging. ACS Chem Neurosci. 2021: 12:2553-4.
 - 151 Caschera L, Lazzara A, Piergallini L, Ricci D, Tuscano B, Vanzulli A. Contrast agents in diagnostic imaging: present and future. Pharmacol Res. 2016; 110:65-75.
 - Cormode DP, Naha PC, Fayad ZA. [6] Nanoparticle contrast agents for computed tomography: a focus on micelles. Contrast Media Mol Imaging. 2014; 9:37–52.
 - Sun Y, Yu M, Liang S, Zhang Y, Li C, Mou T, [7] Yang W, Zhang X, Li B, Huang C, Li F. Fluorine-18 labeled rare-earth nanoparticles for positron emission tomography (PET) imaging of sentinel lymph node. Biomaterials. 2011; 32:2999-3007
 - Ramalho J, Semelka RC, Ramalho M, Nunes [8] RH, AlObaidy M, Castillo M. Gadoliniumbased contrast agent accumulation and toxicity: an update. Am J Neuroradiol. 2016; 37:1192.

- [9] Hille, B. 2001. Ion Channels of Excitable Membranes. Third edition. Sinauer Associates, Inc., Sunderland, MA. 814 pp.
- [10] Ashcroft, F. 2000. Ion Channels and Disease. Academic Press, San Diego, CA. 481 pp.
- [11] Lang F, Stournaras C. Ion channels in cancer: future perspectives and clinical potential. Philos Trans R Soc Lond B Biol Sci 2014; 369: 20130108
- [12] Preussat K, Beetz C, Schrey M, Kraft R, Wolfl S, Kalff R et al. Expression of voltage-gated potassium channels Kv1.3 and Kv1.5 in human gliomas. Neurosci Lett 2003; 346: 33–36
- [13] Comes N, Bielanska J, Vallejo-Gracia A, Serrano-Albarras A, Marruecos L, Gomez D et al. The voltage-dependent K(+) channels Kv1.3 and Kv1.5 in human cancer. Front Physiol 2013; 4: 283
- [14] Pardo LA, Stuhmer W. Eag1: an emerging oncological target. Cancer Res 2008; 68: 1611–1613.
- [15] Downie BR, Sanchez A, Knotgen H, Contreras-Jurado C, Gymnopoulos M, Weber C et al. Eag1 expression interferes with hypoxia and Jou homeostasis and induces angiogenesis in Scientumors. J Biol Chem 2008; 283: 36234–36240.
- Huang X, Dubuc AM, Hashizume R, Berg J, He Y, Wang J et al. Voltage-gated potassium channel EAG2 controls mitotic entry and tumor growth in medulloblastoma via regulating cell volume dynamics. Genes Dev 2012; 26: 1780–1796
- [17] Wang H, Zhang Y, Cao L, Han H, Wang J, Yang B et al. HERG K+ channel, a regulator of tumor cell apoptosis and proliferation. Cancer Res 2002; 62: 4843–4848.
- [18] Chandy KG, Wulff H, Beeton C, Pennington M, Gutman GA, Cahalan MD. K+ channels as targets for specific immunomodulation. Trends Pharmacol Sci 2004; 25: 280–289.
- [19] Pardo LA, Stuhmer W. The roles of K(+) channels in cancer. Nat Rev Cancer 2014; 14: 39–48.
- [20] Liu X, Chang Y, Reinhart PH, Sontheimer H, Chang Y. Cloning and characterization of glioma BK, a novel BK channel isoform highly expressed in human glioma cells. J Neurosci 2002; 1840–1849.
- [21] Grossinger EM, Weiss L, Zierler S, Rebhandl S, Krenn PW, Hinterseer E et al. Targeting

- proliferation of chronic lymphocytic leukemia (CLL) cells through KCa3.1 blockade. Leukemia 2014; 28: 954–958.
- [22] Hammadi M, Chopin V, Matifat F, Dhennin-Duthille I, Chasseraud M, Sevestre H et al. Human ether a-gogo K(+) channel 1 (hEag1) regulates MDA-MB-231 breast cancer cell migration through Orai1-dependent calcium entry. J Cell Physiol 2012; 7: 3837–3846.
- [23] Schwab A, Fabian A, Hanley PJ, Stock C. Role of ion channels and transporters in cell migration. Physiol Rev 2012; 92: 1865–1913.
- [24] Khaitan D, Sankpal UT, Weksler B, Meister EA, Romero IA, Couraud PO et al. Role of KCNMA1 gene in breast cancer invasion and metastasis to brain. BMC Cancer 2009; 9: 258.
- [25] Chantome A, Potier-Cartereau M, Clarysse L, Fromont G, Marionneau-Lambot S, Gueguinou M et al. Pivotal role of the lipid Raft SK3-Orail complex in human cancer cell migration and bone metastases. Cancer Res 2013; 73: 4852–4861.
- [26] Schwab A, Schuricht B, Seeger P, Reinhardt J, Dartsch PC. Migration of transformed renal epithelial cells is regulated by K+ channel modulation of actin cytoskeleton and cell arch and volume. Pflugers Arch 1999; 438: 330–337
 - Ruggieri P, Mangino G, Fioretti B, Catacuzzeno L, Puca R, Ponti D et al. The inhibition of KCa3.1 channels activity reduces cell motility in glioblastoma derived cancer stem cells. PloS One 2012; 7: e47825.
 - [28] Berridge MJ, Bootman MD, Roderick HL. Calcium signalling: dynamics, homeostasis and remodelling. Nat Rev Mol Cell Biol. 2003; 4:517–29.
 - [29] Okada H, Okabe K, Tanaka S. Finely-tuned calcium oscillations in osteoclast differentiation and bone resorption. Int J Mol Sci. 2020; 22:180.
 - [30] Yang Z, Yue Z, Ma X, Xu Z. Calcium homeostasis: a potential vicious cycle of bone metastasis in breast cancers. Front Oncol. 2020; 10:293.
 - [31] Marchi S, Giorgi C, Galluzzi L, Pinton P. Ca2+fluxes and cancer. Mol Cell. 2020; 78:1055–69.
 - [32] Mikoshiba K, Furuichi T, Miyawaki A. Structure and function of IP3 receptors. Semin Cell Biol. 1994; 5:273–81.
 - [33] Roberts-Thomson SJ, Chalmers SB, Monteith GR. The calcium-signaling toolkit in cancer:

- remodeling and targeting. Cold Spring Harb Perspect Biol. 2019; 11:a035204.
- Roberts-Thomson SJ, Chalmers SB, Monteith [34] GR. The calcium-signaling toolkit in cancer: remodeling and targeting. Cold Spring Harb Perspect Biol. 2019; 11:a035204.
- Catterall WA. Voltage-gated calcium channels. [35] Cold Spring Harb Perspect Biol. 2011; 3:a003947.
- [36] He L, Si G, Huang J, Samuel ADT, Perrimon N. Mechanical regulation of stem-cell differentiation by the stretch-activated Piezo channel. Nature. 2018: 555:103-6.
- [37] Moran MM, McAlexander MA, Bíoró T, Szallasi A. Transient receptor potential channels as therapeutic targets. Nat Rev Drug Discov. 2011; 10:601-20.
- H.M. Shepard, P. Jin, D.J. Slamon, Z. Pirot, [38] D.C. Maneval Herceptin Handb. Exp. Pharmacol. (2008), pp. 183-219
- [39] N. Prevarskaya, H. Ouadid-Ahidouch, R. Skryma, Y. Shuba Remodelling of Ca2+ transport in cancer: how it contributes to cancer hallmarks? Philos. Trans. R. Soc. Lond. Ser. Bonal Jo Biol. Sci., 369 (2014), Article 20130097 rend in [49] en M. Feng, D.M. Grice, H.M. Faddy, N. Nguyen,
- [40] R.S. Donadi, C.G. Bologa, A. Karlsson, B. Al-lopmeni Lazikani, A. Hersey, T.I. Oprea, J.P. Overington A comprehensive map of molecular drug targets Nat. Rev. Drug Discov., 16 (2017), pp. 19-34
- G. Shapovalov, A. Ritaine, R. Skryma, N. [41] Prevarskaya Role of TRP ion channels in and tumorigenesis cancer Semin. Immunopathol., 38 (2016), pp. 357-369
- N. Prevarskaya, R. Skryma, Y. Shuba Targeting [42] Ca(2)(+) transport in cancer: close reality or long perspective? Expert Opin. Ther. Targets, 17 (2013), pp. 225-241
- S. Fu, H. Hirte, S. Welch, T.T. Ilenchuk, T. [43] Lutes, C. Rice, N. Fields, A. Nemet, D. Dugourd, S. Piha-Paul, V. Subbiah, L. Liu, J. Gong, D. Hong, J.M. Stewart First-in-human phase I study of SOR-C13, a TRPV6 calcium channel inhibitor, in patients with advanced solid tumors Investig. New Drugs, 35 (2017), pp. 324-333
- [44] V. Lehen'kyi, M. Raphael, N. Prevarskaya. The role of the TRPV6 channel in cancer J. Physiol., 590 (2012), pp. 1369-1376

- [45] A. A. Peters, P.T. Simpson, J.J. Bassett, J.M. Lee, L. Da Silva, L.E. Reid, S. Song, M.O. Parat, S.R. Lakhani, P.A. Kenny, S.J. Roberts-Thomson, G.R. Monteith Calcium channel TRPV6 as a potential therapeutic target in estrogen receptor-negative breast cancer Mol. Cancer Ther., 11 (2012), pp. 2158-2168
- [46] M. Raphael, V. Lehen'kyi, M. Vandenberghe, B. Beck, S. Khalimonchyk, F. Vanden Abeele, L. Farsetti, E. Germain, A. Bokhobza, A. Mihalache, P. Gosset, C. Romanin, P. Clezardin, R. Skryma, N. Prevarskaya TRPV6 calcium channel translocates to the plasma membrane via Orai1-mediated mechanism and controls cancer cell survival Proc. Natl. Acad. Sci. U. S. A., 111 (2014), pp. E3870-E3879
- [47] V. Lehen'kyi, M. Flourakis, R. Skryma, N. Prevarskaya TRPV6 channel controls prostate cancer cell proliferation via Ca(2+)/NFATdependent pathways Oncogene, 26 (2007), pp. 7380-738
 - S. Yang, J.J. Zhang, X.Y. Huang Orai1 and STIM1 are critical for breast tumor cell migration and metastasis Cancer Cell, 15 (2009), pp. 124-134
- R. Santos, O. Ursu, A. Gaulton, A.P. Bento, arch and S. Leitch, Y. Wang, S. Muend, P.A. Kenny, S. Sukuma, J. Roberts-Thomson, G.R. Monteith, R. Rao Store-independent activation of Orai1 by SPCA2 in mammary tumors Cell, 143 (2010), pp. 84-98
 - [50] D. McAndrew, D.M. Grice, A.A. Peters, F.M. Davis, T. Stewart, M. Rice, C.E. Smart, M.A. Brown, P.A. Kenny, S.J. Roberts-Thomson, G.R. Monteith ORAI1-mediated calcium influx in lactation and in breast cancer Mol. Cancer Ther., 10 (2011), pp. 448-460.
 - [51] W.H. Lee, L.Y. Choong, T.H. Jin, N.N. Mon, S. Chong, C.S. Liew, T. Putti, S.Y. Lu, C. Harteneck, Y.P. Lim, TRPV4 plays a role in breast cancer cell migration via Ca (2+)activation dependent of AKT and downregulation of E-cadherin cell cortex protein, Oncogene 6 (2017) e338.
 - [52] GlaxoSmithKline, A Study to Evaluate the Effect of the Transient Receptor Potential Vanilloid 4 (TRPV4) Channel Blocker, GSK2798745, on Pulmonary Gas Transfer and Respiration in Patients With Congestive Heart Failure, US National Library of Medicine, 2017, ClinicalTrials.gov.

[66]

- [53] M.C. Sallan, A. Visa, S. Shaikh, M. Nager, J. Herreros, C. Canti, T-type Ca(2+) channels: T for targetable, Cancer Res. 78 (2018) 603–609.
- [54] C. Tian, L. Du, Y. Zhou, M. Li, Store-operated CRAC channel inhibitors: opportunities and challenges, Future Med. Chem. 8 (2016) 817–83
- [55] Y. Zhang, N. Cruickshanks, F. Yuan, B. Wang, M. Pahuski, J. Wulfkuhle, I. Gallagher, A.F. Koeppel, S. Hatef, C. Papanicolas, J. Lee, E.E. Bar, D. Schiff, S.D. Turner, E.F. Petricoin, L.S. Gray, R. Abounader, Targetable T-type calcium channels drive glioblastoma, Cancer Res. 77 (2017) 3479–3490
- [56] A.S. Ay, N. Benzerdjeb, H. Sevestre, A. Ahidouch, H. Ouadid-Ahidouch, Orai3 constitutes a native store-operated calcium entry that regulates non-small cell lung adenocarcinoma cell proliferation, PLoS One 8 (2013) e72889.
- [57] Y. Shi, X. Ding, Z.H. He, K.C. Zhou, Q. Wang, Y.Z. Wang, Critical role of TRPC6 channels in G2 phase transition and the development of human oesophageal cancer, Gut 58 (2009) 1443–1450.
- [58] S.L. Yang, Q. Cao, K.C. Zhou, Y.J. Feng, Y.Z. Wang, Transient receptor potential channel C3 contributes to the progression of human ovarian cancer, Oncogene 28 (2009) 1320–1328
- [59] A.A. Peters, S.Y.N. Jamaludin, K. Yapa, S. Chalmers, A.P. Wiegmans, H.F. Lim, M.J.G. Milevskiy, I. Azimi, F.M. Davis, K.S. Northwood, E. Pera, D.L. Marcial, E. Dray, N.J. Waterhouse, P.J. Cabot, T.J. Gonda, P.A. Kenny, M.A. Brown, K.K. Khanna, S.J. Roberts-Thomson, G.R. Monteith, Oncosis and apoptosis induction by activation of an overexpressed ion channel in breast cancer cells, Oncogene 36 (2017) 6490–6500.
- [60] L. Zhang, G.J. Barritt, Evidence that TRPM8 is an androgen-dependent Ca2+ channel required for the survival of prostate cancer cells, Cancer Res. 64 (2004) 8365–8373.
- [61] B. Dziegielewska, D.L. Brautigan, J.M. Larner, J. Dziegielewski, T-type Ca2+ channel inhibition induces p53-dependent cell growth arrest and apoptosis through activation of p38-MAPK in colon cancer cells, Mol. Cancer Res. 12 (2014) 348–358
- [62] H. Liu, J.D. Hughes, S. Rollins, B. Chen, E. Perkins, Calcium entry via ORAI1 regulates

- glioblastoma cell proliferation and apoptosis, Exp. Mol. Pathol. 91 (2011) 753–760.
- [63] M. Faouzi, F. Hague, M. Potier, A. Ahidouch, H. Sevestre, H. Ouadid-Ahidouch, Downregulation of Orai3 arrests cell-cycle progression and induces apoptosis in breast cancer cells but not in normal breast epithelial cells, J. Cell. Physiol. 226 (2011) 542–551
- [64] Y. Zhang, N. Cruickshanks, F. Yuan, B. Wang, M. Pahuski, J. Wulfkuhle, I. Gallagher, A.F. Koeppel, S. Hatef, C. Papanicolas, J. Lee, E.E. Bar, D. Schiff, S.D. Turner, E.F. Petricoin, L.S. Gray, R. Abounader, Targetable T-type calcium channels drive glioblastoma, Cancer Res. 77 (2017) 3479–3490.
- [65] X. Ma, Y. Cai, D. He, C. Zou, P. Zhang, C.Y. Lo, Z. Xu, F.L. Chan, S. Yu, Y. Chen, R. Zhu, J. Lei, J. Jin, X. Yao, Transient receptor potential channel TRPC5 is essential for P-glycoprotein induction in drug-resistant cancer cells, Proc. Natl. Acad. Sci. U. S. A. 109 (2012) 16282–16287
 - P. Vacher, A.M. Vacher, R. Pineau, S. Latour, I. Soubeyran, C. Pangault, K. Tarte, P. Soubeyran, T. Ducret, L. Bresson-Bepoldin, Localized store-operated calcium influx represses CD95-dependent apoptotic effects of rituximab in non-Hodgkin B lymphomas, J. Immunol. 195 (2015) 2207–2215
- [67] A.A. Peters, M.J. Milevskiy, W.C. Lee, M.C. Curry, C.E. Smart, J.M. Saunus, L. Reid, L. da Silva, D.L. Marcial, E. Dray, M.A. Brown, S.R. Lakhani, S.J. Roberts Thomson, G.R. Monteith, The calcium pump plasma membrane Ca(2+)-ATPase 2 (PMCA2) regulates breast cancer cell proliferation and sensitivity to doxorubicin, Sci. Rep. 6 (2016) 25505
- [68] M. Nabissi, M.B. Morelli, M. Santoni, G. Santoni, Triggering of the TRPV2 channel by cannabidiol sensitizes glioblastoma cells to cytotoxic chemotherapeutic agents, Carcinogenesis 34 (2013) 48–57
- [69] D. Klumpp, M. Misovic, K. Szteyn, E. Shumilina, J. Rudner, S.M. Huber, Targeting TRPM2 channels impairs radiation-induced cell cycle arrest and fosters cell death of T cell leukemia cells in a Bcl-2-dependent manner, Oxidative Med. Cell. Longev. 2016 (2016) 8026702.
- [70] D. Klumpp, S.C. Frank, L. Klumpp, E.C. Sezgin, M. Eckert, L. Edalat, M. Bastmeyer, D. Zips, P. Ruth, S.M. Huber, TRPM8 is required

- for survival and radioresistance of glioblastoma cells, Oncotarget 8 (2017) 95896-95913.
- A. Tosatto, R. Sommaggio, C. Kummerow, [71] R.B. Bentham, T.S. Blacker, T. Berecz, M.R. Duchen, A. Rosato, I. Bogeski, G. Szabadkai, R. Rizzuto, C. Mammucari, The mitochondrial calcium uniporter regulates breast cancer progression via HIF1alpha, EMBO Mol. Med. 8 (2016) 569-585
- [72] M. Feng, D.M. Grice, H.M. Faddy, N. Nguyen, S. Leitch, Y. Wang, S. Muend, P.A. Kenny, S. S.J. Roberts-Thomson, Sukumar, Monteith, R. Rao, Store-independent activation of Orai1 by SPCA2 in mammary tumors, Cell 143 (2010) 84-98
- I. Frischauf, M. Litvinukova, R. Schober, V. [73] Zayats, B. Svobodova, D. Bonhenry, V. Lunz, S. Cappello, L. Tociu, D. Reha, A. Stallinger, A. Hochreiter, T. Pammer, C. Butorac, M. Muik, K. Groschner, I. Bogeski, R.H. Ettrich, C. Romanin, R. Schindl, Transmembrane helix connectivity in Orail controls two gates for calcium-dependent transcription, Sci. Signal. 10 (2017).
- F. Beuschlein, S. Boulkroun, A. Osswald, T. onal Jo [74] Penton, V.R. Schack, L. Amar, E. Fischer, A. arch and Walther, P. Tauber, T. Schwarzmayr, S. Diener, Johnson E. Graf, B. Allolio, B. Samson-Couterie, A. Benecke, M. Quinkler, F. Fallo, P.F. Plouin, F. Mantero, T. Meitinger, P. Mulatero, X. Jeunemaitre, R. Warth, B. Vilsen, M.C. Zennaro, T.M. Strom, M. Reincke, Somatic mutations in ATP1A1 and ATP2B3 lead to aldosterone producing adenomas and secondary hypertension, Nat. Genet. 45 (2013) 440–444.
- P. Tauber, B. Aichinger, C. Christ, J. Stindl, Y. [75] Rhayem, F. Beuschlein, R. Warth, S. Bandulik, Cellular pathophysiology of an adenoma-associated mutant of the plasma membrane Ca(2+)-ATPase ATP2B3, Endocrinology 157 (2016) 2489-2499.
- [76] P.B. Davis, U. Yasothan, P. Kirkpatrick, Ivacaftor, Nat. Rev. Drug Discov. 11 (2012) 349-350.
- Rao et al., Voltage-Gated Ion Channels in [77] Cancer Cell Proliferation. Cancers (Basel). 2015; 7(2): 849-875.

- [78] Schwab and Stock, Ion channels and transporters in tumor cell migration and invasion. Philos Trans R Soc Lond B Biol Sci. 2014; 369(1638): 20130102.
- Mycielska et al., Expression of Na+-dependent [79] citrate transport in a strongly metastatic human prostate cancer PC-3M cell line: regulation by voltage-gated Na+ channel activity. *J Physiol*. 2005; 563(Pt 2):393-408.
- [80] Krasowska et al., Patterning of endocytic vesicles and its control by voltage-gated Na+ channel activity in rat prostate cancer cells: fractal analyses. Eur Biophys J. 2004; 33(6):535-42
- [81] Bortner and Cidlowski, Ion channels and apoptosis in cancer. Philos Trans R Soc Lond B Biol Sci. 2014; 369(1638): 20130104
- [82] Jang S, et al., Kv1.3 voltage-gated K+ channel subunit as a potential diagnostic marker and therapeutic target for breast cancer. BMB Rep. 2009; 42:535–9
- [83] Hemmerlein B. et al., Overexpression of EAG1 potassium channels in clinical tumors. Mol Cancer 2006; 5: 41
- Wieland, H.N. Nielsen, U.D. Lichtenauer, D. in [84] an Abdul M, et al., Reduced Kv1.3 Potassium channel expression in human prostate cancer. J Membrane Biol. 2006; 214:99-102
 - [85] Fraser J et al., Contribution of functional voltage-gated Na+ channel expression to cell behaviors involved in the metastatic cascade in rat prostate cancer: I. Lateral motility. Physiol. 2003; 195(3):479-87
 - Nelson et al., Nav1.5 regulates breast tumor [86] growth and metastatic dissemination in vivo. Oncotarget 2015; 6(32):32914-29
 - [87] Fraser J et al., Voltage-gated sodium channel expression and potentiation of human breast cancer metastasis. Clinical Cancer Research, 2005; 11(15)
 - [88] Chen, R. et al. (2014). Cav1.3 channel a1D protein is overexpressed and modulates androgen receptor transactivation in prostate cancers. Urol. Oncol. Semin. Orig. Investig. 2014; 32, 524–536
 - [89] Wang, CY. et al., Meta-analysis of public microarray datasets reveals voltage-gated calcium gene signatures in clinical cancer patients. PloS One. 2015a; 10